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Molecular Profiling of Activated Neurons by Phosphorylated Ribosome Capture

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SUMMARY

The mammalian brain is composed of thousands of interacting neural cell types. Systematic approaches to establish the molecular identity of functional populations of neurons would advance our understanding of neural mechanisms controlling behavior. Here, we show that ribosomal protein S6, a structural component of the ribosome, becomes phosphorylated in neurons activated by a wide range of stimuli. We show that these phosphorylated ribosomes can be captured from mouse brain homogenates, thereby enriching directly for the mRNAs expressed in discrete subpopulations of activated cells. We use this approach to identify neurons in the hypothalamus regulated by changes in salt balance or food availability. We show that galanin neurons are activated by fasting and that prodynorphin neurons restrain food intake during scheduled feeding. These studies identify elements of the neural circuit that controls food intake and illustrate how the activity-dependent capture of cell-type-specific transcripts can elucidate the functional organization of a complex tissue.

INTRODUCTION

A basic goal of neuroscience is to link the activity of specific neuronal cell types to the various functions of the brain. This task is complicated by the extraordinary cellular diversity of the

The ability to profile the genes uniquely expressed that respond to a stimulus would facilitate the systematic identification of the cell types that control behavior. Molecular identification of these cells would also enable their manipulation *in vivo* by using technologies that are possible to activate or inhibit neurons with light (Yamamoto et al., 2011), generate transcriptional profiles from neurons using fluorescently tagged ribosomes (Heiman et al., 2008; Sanz et al., 2011), and label and record from neurons by using fluorescently tagged ribosomes (Gong et al., 2003). These tools achieve their selectivity by targeting protein expression using a promoter from a cell-type-specific marker gene, but in many cases, the marker genes for specific functional populations of neurons are unknown (Gong et al., 2007).

Immediate early genes such as *c-fos* have been used to visualize the neurons that respond to numerous stimuli (Curran and Palmiter, 1991). However, despite its utility in marking neurons that have been biochemically activated, *c-fos* staining reveals the genetic identity of the labeled cells. Characterization of the genetic identity of the labeled cells requires coexpression of an activation marker such as *c-fos* with a limited set of candidate genes requires processing large numbers of histologic sections (Isogai et al., 2011). For this reason, alternative methods are needed to profile gene expression in discrete subpopulations of activated neurons in the brain.

Here, we show that phosphorylation of the ribosome can be used as a molecular tag to retrieve RNA selectively from activated neurons. This enables the unbiased discovery of genes that are uniquely expressed in a functional population of neurons. By quantifying in parallel the enrichment of specific markers, it is possible to assess the activation or repression of numerous cell types in a complex tissue, revealing the coordinated regulation of ensembles of neurons in response to a stimulus.